Centre for Biological Engineering

RISK ASSESSMENT REVIEW/REVISION RECORD

Risk Assessment Ref No:	CBE/BRA/008	Version Number

This risk assessment should be reviewed **annually** or more frequently if there is any change in the work, or if new information becomes available that indicates the assessment may no longer be valid. **This form should be attached to the front of the current version of the risk assessment or to the new version of the risk assessment if one is issued**

The following review/revision has been carried out on the dates indicated and either the assessment remains valid or it has been amended as indicated.	
Name(s) of reviewer: A. Chandra	Date: 11 May 2010
Signature: A. Chandra	
Amendments: This biological risk assessment is withdrawn on 11 May 2010. All work on Grand Challenge Remedi WP 3.2 has ceased.	
This review or revision must be approved by the person supervising the work and the CBE Quality Manager. Significant changes may require a revised version of the risk assessment to be issued for re-approval by the local BGMSA and/ or the BSO and/or GM Safety Committee, as appropriate.	
Name of Approver: P. Hourd	Date: 11 May 2010
Position: QM	
Signature: P Hourd	

CBE/BRA/008 rev 2

Centre for Biological Engineering

RISK ASSESSMENT REVIEW RECORD

Risk Assessment Ref No:	REMEDI WP3(ii) Project (no assessment number issued)

This risk assessment should be reviewed **annually** or more frequently if there is any change in the work, or if new information becomes available that indicates the assessment may no longer be valid. Reviews have been carried out on the following dates and either the assessment remains valid or it has been amended as indicated. This record should be attached to the amended risk assessment where appropriate.

Name of reviewer: Paul Hourd	Date: 11th May 2009
Signature:	

Amendments:

The above risk assessment has been reviewed following the proposed transfer of work activities involving biological agents within the Remedi WP3(ii) Project to a new Containment Level 2 facility in the Centre for Biological Engineering (CBE) at Holywell Park. The CBE facility contains a self-contained, Containment Level 2 Laboratory Unit comprising 7 laboratories with ancillary rooms such as changing rooms, store rooms and an autoclave room. The CBE Laboratory Unit is a shared multi-user facility. The primary purpose of the Unit is translational research aimed at the generation of new medical therapies, healthcare technologies and associated enabling technologies with a particular focus on manufacturing and bioprocessing. Much of the work in the Unit involves biological material. The Unit has therefore been designed as a controlled environment and operates under a Quality Management System to both be compliant to the necessary regulations, to ensure research quality and relevance and to protect research materials.

This risk assessment has therefore been amended to incorporate the requirements of a new local Code of Practice and a new Quality Management System that have been drawn up (restricted access available at https://internal.lboro.ac.uk/restricted/wolfson/Healthcare_SOP/) to ensure that Containment Level 2 work within the CBE Laboratory Unit is compliant with the 2002 COSHH (amended) Regulations and the Loughborough University Biological Safety Policy.

These amendments, **highlighted in yellow**, record the minor changes to working practice (documented in revised SOPs), that are necessary to ensure that existing precautions and control measures are adequately transferred to the new facility and to ensure that there are no additional risks to laboratory personnel, workers in the entire CBE, people in the external environment or to the environment itself. Since there are no significant changes to the biological hazards (in relation to the biological agents used) or nature of the work, this risk assessment is still relevant to the work activity of the project.

CBE/BRA/008

Loughborough University

For Health and Safety Unit Use only
ASSESSMENT NO.
REMEDI WP3 ii
31/10/08

RISK ASSESSMENT OF WORK WITH BIOLOGICAL MATERIALS

Please note the following before completing this form:

1. University Health and Safety Policy requires that risk assessment of all work with biological materials must be carried out in advance of work commencing. A key requirement of The Control of Substances Hazardous to Health Regulations (COSHH) is to assess the risks associated with any work activity involving the use of biological materials that may contain biological agents.
2. YOU SHOULD COMPLETE ALL OF PART A, THE APPROPRIATE SECTION(S) OF PART B, AND ALL OF PART C. THIS FORM SHOULD BE SUBMITTED TO THE HEALTH AND SAFETY UNIT FOR REVIEW (VIA YOUR DEPARTMENTAL SAFETY OFFICER)
3. It is the responsibility of the Principal Investigator to ensure compliance to these requirements and that this risk assessment remains valid.
4. This risk assessment form **IS NOT** for assessing the risks associated with **Genetically Modified Organism activities**.

Date Submitted:	31st October 2008	Date Approved:	9th November 2008

PART A: Please provide the following general information:

School/Department			
Healthcare Engineering, Centre for Biological Engineering (CBE)			
The Project			
Title of Project: Remedi Grand Challenge: Regenerative Medicine - a new industry This Risk Assessment applies to the final one and a half years Work Package 3(ii) of the aforementioned project			
Project Reference Number: Remedi Application Reference: EP/C534247/1. Project Reference: REMEDI/WP3(ii)			
Person responsible for this work (*Principle Investigator*):			
Name: David Williams:		Position: Professor of Healthcare Engineering	
Department: Healthcare Engineering		University School: Wolfson School of Mechanical and Manufacturing Engineering/ CBE	
Person conducting this assessment			
Name: Carolyn Thomas/Paul Hourd		Position: Laboratory Manager/Remedi Project Manager	
Department:	Healthcare Engineering	Date Risk Assessment Undertaken:	October 2008
Proposed Project Start Date:	Nov 2008	Proposed Project End Date:	28.02.2010

Assessment Review:
required at least once a year or immediately following any significant change to the project

	Review 1	Review 2	Review 3	Review 4	Review 5
Due Date	Oct 09				
Date Conducted	23 April 09				

A1 PROJECT SUMMARY

A1.1 Scientific Goals of the Project *Brief yet clear outline only*

Work Package 3(ii) comprises a connected programme of work (CPW) consisting of series of integrated and coherent studies connected as part of a continuing remedi programme of work. The CPW is focussed on process discovery, optimisation and continuous improvement for the manufacture of human cellular feedstocks. The goal is to demonstrate the transfer of manual, bench-scale human cell culture processes to scaled automated processes using a commercially available automated platform (ConpacT SelecT). Each component part of the project aims to provide automated demonstrators with measured Critical to Quality (CTQ) process output verified as equivalent to the benchmark manual process.

A1.2 Description of the Experimental Procedures

Describe laboratory procedures to be used and highlight any non-standard laboratory operations

1) Human osteoblast-like cell line (HOS) – Seeded onto tissue culture treated plastic (e.g. T75 flasks) and cultured at 37°C 5% CO^2 until 70% confluent, followed by trypsinisation and subsequent culture. Manual and automated (T175 flasks) cell culture techniques will be used.
2) Human mesenchymal progenitor cells (hMSCs)- Isolated via centrifugation of fresh bone marrow aspirate over a defined a defined density cell separation product (Histopaque™). Cells will be seeded onto tissue culture treated plastic (e.g. T175 flasks) and cultured at 37°C 5% CO^2 until 70% confluent, followed by trypsinisation and subsequent subculture .Manual and automated cell culture techniques will be used . Cryopreservation and subsequent thawing and subculture will be conducted at various points in the process.
3) Human mesenchymal stem cells from Bone Marrow mononuclear cells (hMSCs) - Cells will be seeded onto tissue culture treated plastic (e.g. T175 flasks) and cultured at 37°C 5% CO^2 until 70% confluent, followed by trypsinisation and subsequent subculture. Manual and automated cell culture techniques will be used. Cryopreservation and subsequent thawing and subculture will be conducted at various points in the process.
4)Endothelial Progenitor from peripheral blood mononuclear cells- (derived from Poietics® mononuclear cells from Human G-CSF Mobilised cells), Cells will be cultured on human fibronectin coated T175 flasks and the adherent cells passaged according to the general culture method.
5) Human Embryonic cell line (hES)-(Hues-1, Hues-9) - hES cells will be cultured in treated plastic vessels (e.g. T175 flasks) and cultured at 37°C 5% CO^2, followed by enzyme digestion and subsequent subculture. Cryopreservation and subsequent thawing and subculture will be conducted at various points in the process. Manual and automated cell culture techniques will be used.
6) Mouse Embryonic Fibroblasts (MEFs) – MEF cells obtained in 15 T175 flasks from the University of Nottingham will be cultured at 37°C 5% CO_2. Media in the cells will be changed every 24h hours, pooled and frozen in 100ml Nalgene sterile media bottles for subsequent hES culture. Various manual and automated culture routines will be used.

All procedures will be conducted in accordance with the laboratory Quality Management System (QMS) requirements, Good Cell Culture Practice, Good Aseptic Technique, the local Code of Practice (COP) and the Loughborough University Biological Safety Policy.

PART B: Please provide information in one or more of the following sections, as appropriate. Only sections which you complete should be submitted:

Section 1: *micro-organisms (prions, viruses, bacteria, fungi, parasites in ACDP category 2 and pathogens controlled by the Department for the Environment, Food and Rural Affairs).* ***[Work with ACDP category 3 and 4 pathogens is not currently permitted in the University.]***

Section 2: *cell cultures, tissues, blood, body fluids or excreta*

Section 3: *plants and plant material*

Section 4: *animals and animal tissues*

SECTION 2: CELL CULTURES, TISSUES, BLOOD, BODY FLUIDS OR EXCRETA

B2.1 HAZARD AND RISK IDENTIFICATION: NATURE OF CELLS, TISSUES OR BODY FLUIDS

*This information gives an indication of the **potential** harm that the biological material may cause*

B2.1.1 List all cells or tissues to be used. *For cells indicate if primary, continuous or finite.*

Indicate in the adjacent box if Not Relevant (N/R)			
Cell or tissue type and ID	Organ Source	Species	From where will it be obtained?
1) Human Osteoblast like cell line (HOS)	Bone	Human	European Collection of Cell Culture (ECACC), UK Continuous cell line
2) Human Mesenchymal progenitor cells (hMSCs)	Bone Marrow	Human	Cambrex,UK Primary cell line
3) Human Mesenchymal stem cells from Bone marrow mononuclear cells	Bone Marrow	Human	Lonza,UK Primary Cell line
4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF mobilised cells	Blood	Human	Lonza,UK Primary Cell line
5) Human Embryonic cell line (hES) (Hues-1,)	Embryo	Human	Harvard University,USA Continuous cell line
6) Human Embryonic cell line (hES) (Hues-9)	Embryo	Human	Harvard University,USA Continuous cell line
7) Mouse Embryonic fibroblasts (MEFs)	Skin	Mouse	Nottingham University,UK Continuous cell line

B2.1.2 List all blood, body fluids or excreta to be used

Indicate in the adjacent box if not relevant (N/R			N/R
Material type and ID	Organ Source	Species	From where will it be obtained?

B2.1.3 Has any material listed in section B2.1.1 been genetically modified in any way?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If Yes, complete Genetically Modified Organisms (GMO) Risk Assessment Form	

B2.1.4 Will material be screened for infectious agents (if from a cell culture collection answer B2.1.6)?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	YES

If Yes, provide details of the types of screening and agents screened for:

Cell or tissue type	Screened for
1) Human osteoblast-like cell line (HOS)	See section B2.1.6. Cultured cells in-house tested for mycoplasma
2) Human Mesenchymal progenitor cells (hMSCs)	Basic chemistry panel, complete blood count, HIV-I, Hepatitis B and C, PRP, Urinalysis, sickle cell solubility test and pregnancy test of donors by the supplier. Refer to Cambrex product data sheet*
3) Human Mesenchymal stem cells from bone marrow mononuclear cells	HIV-I, Hepatitis B and C testing by supplier. Refer to Lonza product data sheet*
4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF Mobilised cells	Each donor is tested and found non-reactive by an FDA approved method for the presence of HIV-1, Hepatitis B Virus and Hepatitis C Virus by the supplier. Refer to Lonza Material Safety Data Sheet*. Cultured cells in-house tested for mycoplasma
5) Human Embryonic cell line (hES) (Hues-1,)	Academic source. No material safety data sheet available See B2.1.6
6) Human Embryonic cell line (hES) (Hues-9,)	Academic source. No material safety data sheet available See B2.1.6
7) Mouse Embryonic Fibroblasts (MEFs)	Academic source. No material safety data sheet available

** (All Material Safety Data Sheets will be held in a folder in the Centre for Biological Engineering Research Office)*

B2.1.5 Will any clinical history (if relevant) be provided with this material?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	Yes

If yes give details:

(2) Human Mesenchymal progenitor cells (hMSCs) – A data sheet is provided with the cells containing information on sample collection and clinical history of donors.

(4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF Mobilised cells - A data sheet is provided with the cells containing information on sample collection and clinical history of donors.

If yes, will a policy of rejection of samples from diseased patients be adopted? Explain

Not relevant for materials (2) and (4) because diseased patient samples are not procured by the commercial supplier/vendor. The data sheets provide information on donor eligibility criteria.

If yes, how will the information be disseminated in the course of the project?

Not relevant for materials (2) and (4) – donor eligibility is determined by pre-screening tests carried out by the supplier as above.

If yes, will this information be anonomised?

For materials (2) and (4), donor information received from the commercial supplier/vendor is anonymised

B2.1.6 If obtained from a cell culture collection, is safety information provided?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	Yes

If Yes, summarise here:

Two of the agents listed in B2.1.4 are obtained from a Cell Culture Collection

1) Human osteoblast-like cell line – There is no evidence for the presence of infections, virus or toxic products. However, ECACC recommends that cultures are handled in Category 2 containment. Refer to ECACC material data sheet.

2) Human embryonic cell line (hES - Hues-1, Hues-9) – Obtained from the Howard Hughes Medical Institute, Harvard University (originator) solely for internal academic research purposes. The material is experimental in nature and may have hazardous properties since not all of its characteristics are known. See reference Material Use Licence F191, Harvard University office of Technology Development.

B2.2 RISK TO HUMANS

B2.2.1 What is the likelihood of infection of this material? Indicate as None, Low Risk, Medium Risk, High Risk, Known Infected*

Cell type and ID	Risk Category	Justification for Selection
1) Human osteoblast-like cell line (HOS)	Low	Well authenticated/characterised continuous cell line from a culture collection. Has documented provenance and screened for the most serious human pathogens. Screened as described in section B2.1.6. Hazard Group 1 requiring baseline containment level CL1
2)Human Mesenchymal progenitor cell (hMSCs)	Low	Well authenticated/characterised primary cell line from a commercial source. Has documented provenance and screened for the most serious human pathogens. Screened as described in section B2.1.4. Hazard Group 2 requiring baseline containment level CL2
3) Human Mesenchymal stem cell from Bone marrow mononuclear cell.(hMSCs)	Low	Well authenticated/characterised primary cell line from a commercial source. Has documented provenance and screened for the most serious human pathogens. Screened as described in section B2.1.4. Hazard Group 2 requiring baseline containment level CL2
4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF mobilised cells	Low	Well authenticated/characterised primary cell line from a commercial source culture collection. Has documented provenance and screened for the most serious human pathogens. Screened as described in section B2.1.4. Hazard Group 2 requiring baseline containment level CL2
5) Human embryonic cell (hES) (Hues-1,)	Low	Well authenticated continuous cell line obtained from the originators culture collection The cells are not fully characterised but have been utilised extensively in peer reviewed academic research. These cells are sourced from a leading academic laboratory that has successfully deposited cells to the National Institute for Biological Standards and Control stem cell bank. Since these cell lines have been subject to extensive sub-culture the risk of pathogenic agent contamination is very low. Hazard Group 2 requiring baseline containment level CL2
6) Human embryonic cell (hES) (Hues-9)	Low	Well authenticated continuous cell line obtained from the originators culture collection The cells are not fully characterised but have been utilised extensively in peer reviewed academic research. These cells are sourced from a leading academic laboratory that has successfully deposited cells to the National Institute for Biological Standards and Control stem cell bank. Since these cell lines have been subject to extensive sub-culture the risk of pathogenic agent contamination is very low. Hazard Group 2 requiring baseline containment level CL2

7) Mouse Embryonic Fibroblasts (MEFs)	Low	Not fully characterised continuous cell line of animal origin obtained from the academic originator, as described in section B2.1.4. Hazard Group 1 requiring baseline containment level CL1
		If low risk or none proceed to section B2.2.4

see The Managing the risks in laboratories and healthcare premises – available at
http://www.hse.gov.uk/biosafety/biologagents.pdf

B2.2.2 If medium or high risk (section B2.2.1), name and classify the Biological Agents this material could be infected with. List the biological agents and indicate the ACDP hazard group classification*

Name of Agent	Classification

see The Approved List of Biological Agents – available on the Health & Safety website or
http://www.hse.gov.uk/pubns/misc208.pdf.

B2.2.3 Describe the routes of infection (in humans) for these adventitious agents (place a 'X' in the relevant box)

Cell or Tissue type	Percutaneous	Mucocutaneous	Inhalation	Ingestion
Details:				

B2.2.4 Are there any other biological hazards (other than adventitious infectious risk) associated with the materials e.g. tumourogenic cells

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If Yes, describe:	

B2.3 HUMANS AT INCREASED RISK OF INFECTION

B2.3.1 Do any of the agents listed in section 2.1 present an overt risk to humans at increased risk (including immunocompromised workers, pregnant workers, breast feeding mothers)?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, Occupational Health must be consulted:	

B2.4. PROPAGATION OR CONCENTRATION OF ADVENTITIOUS AGENTS

B2.4.1 Will any culturing of this material take place?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	Yes
If yes, identify the cells and the conditions these will grow: All cells listed are cultured using manual and automated techniques (see section A1.2) in tissue culture flasks with liquid cell culture media in a 37 degree Celsius humidified incubator.	

B2.4.2 If culturing, will CD4+ cells be present. Describe what cells and for how long these cultures will be allowed to grow

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, explain:	

B2.4.3 If culturing, what is the maximum volume of culture grown?

Indicate in the adjacent box if Not Relevant (N/R)	
Per Flask	Per experiment
1) Human osteoblast-like cell line (HOS) - $6x10^6$ (75ml/flask)	Variable- depends on experiment
2) Human Mesenchymal progenitor cells (hMSCs) -~10 million (40ml/flask)	~ 200 million (20 flasks)
3) Human Mesenchymal stem cell from Bone Marrow mononuclear cells (hMSCs) - ~10 million (40ml/flask)	~ 200 million (20 flasks)
4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF mobilised cells - ~10 million (40ml/flask)	~ 200 million (20 flasks)
5) Human embryonic cell line (hES – Hues-1) - ~10 million (40ml/flask)	~ 200 million (20 flasks)
6) Human embryonic cell line (hES – Hues-9) - ~10 million (40ml/flask)	~ 200 million (20 flasks)
7) Mouse Embryonic Fibroblasts (MEFs) - ~10 million (40ml/flask)	~ 200 million (20 flasks)

B2.4.4 Will the cells be manipulated in any way that could result in a concentration of any adventitious biological agent present?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, explain:	

B2.5 WORKING WITH MATERIAL DONATED BY YOURSELF OR COLLEAGUES : Persons MUST NOT work with their own cells.

B2.5.1 Will any cells be donated by persons working in or has access to the lab?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, explain what precautions are to be taken to prevent that person being exposed to the cells:	
If yes, where will this material be collected:	
If yes, provide justification for not using a safer source:	
If yes, how will confidentiality be assured:	
If yes, has Ethics Committee approval been obtained:	

B2.6 ENVIRONMENTAL CONSIDERATIONS:

B2.6.1 Are any of the agents capable of causing disease or other harm in animals, fish or plants?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, describe:	

B2.6.2 Will there be any other environmental risks?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, describe:	

B2.7 OTHER HAZARDS

B2.7.1 Are there any other hazards associated with this work? For example, hazardous chemicals, cryogenic gases, ionising radiation.

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	Yes
If yes, identify these: 1) Cryogenic processing with Liquid Nitrogen 2) Trypan Blue for generic cell viability testing 3) Generic and specific hazardous non-biological materials eg Histopaque etc.	
If yes, have these been risk assessed and any necessary approval obtained? 1) Liquid Nitrogen. - Procedures will be carried out by trained personnel in accordance with SOP013 "Use and Maintenance of Liquid Nitrogen Stores". Risk Assessment Reference: SAF/MM/1638 (amended) 2) Trypan Blue - Procedures will be carried out by trained personnel in accordance with SOP029 " Safe Handling and disposal of Trypan Blue". Risk (COSHH) Assessment Reference SAF/MM/1745 3) All Hazardous non-biological materials used in this project eg Trypan Blue; Histopaque etc are subjected to COSHH assessment.	

SECTION 4: ANIMALS AND ANIMAL TISSUES

B4.1 HAZARD AND RISK IDENTIFICATION: NATURE OF ANIMALS OR TISSUE

*This information gives an indication of the **potential** harm that the biological material may cause*

B4.1.1 List all animals or animal tissues to be used

Species	Sex	Source	Anatomical Site	Origin or geographical source
Foetal Calf Serum (FCS)	Unknown	Bovine Foetus	Foetus	Commercial supplier: Cambrex, UK. Sourced from South America according to data sheet
Matrigel ™ (mouse)	Unknown	Engelbreth-Holm-Swarm mouse Sarcoma	Basement Membrane matrix mouse tumour	Commercial supplier: Becton Dickinson, (BD), Oxford, UK

B4.1.2 Is the animal or tissue/body fluid to be worked with infected or to be infected?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No*

If Yes, complete Section 1 of this form

NOTE: FCS product contains material of animal origin. The material contains no hazardous or toxic substances. The material is supplied by a commercial company and sent through the post in secure packaging. Material is liquid and stored frozen at -20C.

NOTE: Matrigel product is derived from mouse colonies that are routinely screened for pathogens. It is tested and found negative for bacteria, fungi and mycoplasma by the supplier. The material is supplied by a commercial company and sent through the post in secure packaging. Material is liquid and stored frozen at -20C.

B4.1.3 Is a carcinogen, drug or other substance to be administered to the animal(s) or present in the tissue?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R

If Yes, complete the appropriate Chemical COSHH Assessment

B4.1.4 Have the investigators that will be performing the work on animals obtained the appropriate Home Office Licence?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R

If No, consult the H&S Office.

B4.1.5 Have Standard Operating Procedures (SOPs) for the proposed work been approved?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R

If No, consult the H&S Office. If Yes attach the signed approval.

B4.2 RISK TO HUMANS

B4.2.1 The disease(s) caused to humans
Describe the type and severity of effects or disease(s) on human health (including infection, allergy, bites and scratches)

Name of animal/animal tissue	Type	Severity
1. Foetal Calf Serum	Under EC regulations (Directive 1999/45/EC), contains no hazardous or toxic substances according to the supplier. Likelihood that it contains substances hazardous to health is low. Animal proteins may be a potential contact irritant	Potential contact irritant
2. Matrigel ™	Product contains no hazardous constituents, or concentrations of all chemical constituents are below the regulatory threshold limits described by Occupational Safety Health Hazard Communication Standard 29CFR 1910.1200 and the EU Directive 91/155/EEC and 93/112/EC. Refer to MSDS*	N/R

B4.2.2 What is the likelihood of infection of this material? INDICATE as None, Low Risk, Medium Risk, High Risk, Known Infected

Name of agent	Risk Category	Justification for Selection
1. Non categorised (refer to MSDS)	None.	FCS product contains no hazardous or toxic substances that require Cambrex to distribute a MSDS according to EC regulations. Well authenticated/characterised product from commercial source. Baseline containment level CL1
2. Non categorised (refer to MSDS)	None	Matrigel™ is used in small quantities for research use only. Well authenticated/characterised product from commercial source. Baseline containment level CL1. Mouse colonies are routinely screened for pathogens via Mouse antibody Production (MAP). Tested and found negative for bacteria, fungi and mycoplasma by the supplier. Refer to Matrigel™ Material Safety Data Sheet*
		If none proceed to section B4.3

B4.2.3 Describe the routes that the effects described in section B4.2.1 are transmitted (place a 'X' in the relevant box)

Percutaneous	Mucocutaneous	Inhalation	Ingestion	N/R
Details:				

B4.3 HUMANS AT INCREASED RISK OF INFECTION

B4.3.1 Do any of the agents listed in section B4.1 present an overt risk to humans at increased risk (including immunocompromised workers, pregnant workers, breast feeding mothers, workers repeatedly handling or multiply dosing animals)?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If yes, Occupational Health must be consulted:	

B4.4. PROPAGATION OR CONCENTRATION OF ADVENTITIOUS AGENTS

B4.4.1 Will any culturing of this material take place?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If yes, complete Section 2 of this form:	

B4.4.2 How many animals will be used?

Indicate in the adjacent box if Not Relevant (N/R)	N/R

B4.5 ENVIRONMENTAL CONSIDERATIONS: Risk to other animals

B4.5.1 Will there be any risk other animals?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe:	

B4.5.2 Will there be any other environmental risks?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If yes, describe:	

PART C: CONTROL MEASURES

C1. CONTROL MEASURES

The risk of exposure must be prevented or adequately controlled to minimise the chance of harm arising. COSHH Regulations require minimum containment measures for laboratories handling organisms from the different ACDP hazard groups (http://www.hse.gov.uk/pubns/misc208.pdf)
The hazard group number typically indicates the level of containment (includes physical measures & working practices) that must be used for its handling).

C1.1 Preventing Exposure

C1.1.1 Substitution with a Safer Alternative

Is substitution with a safer alternative practical, by for example, replacement of a clinical strain or pathogen with one that is lab adapted? Provide reasons for your answer:

Cell /Tissue Type	Substitution available?
1) Human osteoblast-like cell line (HOS)	Not available. Use of these specific human cells is critical to the value of the research. Cells are sourced from established commercial suppliers according to SOP048 " Generation of Risk Assessments for New Materials and Processes" and SOP036, "Maintenance of a Quality Laboratory Environment"
2 Human Mesenchymal progenitor cell (hMSCs)	
3) Human Mesenchymal stem cells from bone marrow mononuclear cells (hMSCs) -	
4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF Mobilised cells	
5) Human Embroyonic cell line (Hues -1)	Not available. Use of these specific embryonic cells is critical to the value of the research. Cells are sourced from an established culture collection at Harvard University.
6) Human Embroyonic cell line (Hues -9)	
7) Mouse Embryonic fibroblasts (MEFs)	Not available. Use of these specific cells is critical to the value of the research. These cells are sourced from Nottingham University and their specific use designated by the transferred protocol for eSC culture.
8) Foetal Calf Serum (FCS)	Not available. Use is needed for supplementing media for cell culture. Sourced from established commercial supplier according to SOP048 " Generation of Risk Assessments for New Materials and Processes" and SOP036, "Maintenance of a Quality Laboratory Environment"
9) Matrigel™	Not Available. Use designated by specific transferred culture protocols. Sourced from established commercial suppliers.

C1.1.2 Isolation/Segregation

(i) Is/Are the laboratory(s) to be used for this work to be shared with other workers not directly involved in this activity?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes

If yes, provide details:

Access to the Containment Level 2 CBE Laboratory Unit is restricted to authorised laboratory workers with appropriate training in accordance with documented local Code of Practice (COP) and Quality Management System (QMS) requirements for Containment Level 2 work activities involving biological materials (CL2).

The laboratories are locked at all times outside of normal working hours to ensure safe storage of biological agents and unauthorised entry. Keys to the laboratories are only issued to authorised users. Access is also restricted to the building (swipe card) and CBE (key entrance) during normal working hours. Out of Hours/Lone working is logged and permitted subject to risk assessment.

No cleaning personnel are permitted in the CBE Laboratory Unit. Access by other Non-Laboratory or maintenance personnel is subject to risk assessment and Permit-to-Work system documented in the local COP.

(ii) Is access to the laboratory(s) to be used for this work restricted?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes

If yes, provide details:

Access is restricted to people with documented training (authorised access documented in each individual's training record) in accordance with the COP and QMS.

C1.2 Controlling Exposure

C1.2.1 Are sharps (needles, blades, scissors, forceps, glass or capillary tubes) to be used at any stage during this activity?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No

If yes, list the sharps:

The sharps used that may cause damage to the skin include glass microscope slides and cover slips.

If yes, justify there use – is there an alternative?

It is local practice in the CBE laboratory unit that the use of sharps is avoided wherever possible. Glass items are replaced with plastic alternatives where possible. However, the above sharps are essential for microscopy work (according to SOP033; "Use and Maintenance of Haemocytometer" and SOP022; "Use and Maintenance of the Olympus CKX41 Inverted Microscope).

If yes, describe there use and disposal:

Used sharps are placed directly into a sharps containers conforming to BS 7320. Sharps bins are removed when three quarters full and contents rendered rendered safe by autoclaving prior to their removal from site.

If yes, describe any additional precautions employed to reduce risk:

Accident procedures for sharps and glass injuries are displayed in posters in all labs within the Unit

C1.2.2 Containment and Ventilation

(i) Is the use of BSC required for the protection of the worker ie do the work procedures generate aerosols or splashes that pose a risk to workers?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes

If yes, specify the type(s) and when they will be used:

A Class II Biological Safety Cabinet will be used for all manipulations that may produce aerosols or splashes of both Hazard Group (HG) 1 and 2 biological agents according to the following SOPs

1) SOP009, "Use and Maintenance of HERASAFE KS Class II BSC"
2) SOP052, "Use and Maintenance of Bioquell Advanced Microflow Class II Biosafety Cabinet"
3) SOP035, "Use and Maintenance of CompacT SelecT"

This control measure is specifically to protect the worker (when handling HG2 BAs) and ensure protection of research materials as part of a quality assurance discipline (when handling both HG1 and 2 BAs).

The Compact Select is used to seed, feed, maintain, expand and harvest human cells (primary and cell lines) cultured in bar-coded T175 conventional flasks. Both multiple individual flask cultures and multiple flask batches are maintained concurrently and processed without cross contamination, mix-ups or other detrimental effects. The Compact Select is the equivalent of a BSC and incubator in one unit, therefore containing any harmful aerosols and splashes inside the equipment; see section C2.2. Risk Assessment for the Compact Select SAF/MM/1956

(ii) Are there any requirements for room ventilation e.g. negative pressure, temperature control?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No

If yes, specify:

C1.2.3 Transport and Storage within the laboratory

How and where are materials to be stored?

All cell lines/tissues listed in B2.1.1 will be stored in a cryobank or temporary storage in designated cell culture incubators according to the following SOPs :

1) SOP005, "Storage and Transport of Biological Materials"
2) SOP008, "Receipt of Hazardous Biological Material"
3) SOP013, "Use and Maintenance of Liquid Nitrogen Stores"
4) SOP017, "Use and Maintenance of the Galaxy-R Incubator"
5) SOP031, "Cryopreservation and Storage of Mammalian Cell Lines"
6) SOP053, "Use and Maintenance of the Sanyo CO2 Incubator

Foetal Calf Serum and Matrigel will be stored in Fridges and Freezers according to the following SOP's:

1) SOP016 " Use and Maintenance of Fridges and Freezers"
2) SOP005 " Storage and Transport of Biological Material "
3) SOP039 " Storage, Handling and Disposal of Chemicals"

Storage units are located in Laboratories H22 and H23 of the CBE Laboratory Unit

How will this material be transported within the laboratory e.g. between BSC and incubator? Detail the containment measures which will be used to prevent or contain accidental splashes or spills.

Cells will always be transferred in closed secondary containers large enough to carry the designated material. . Appropriate spill response procedures are posted in the lab and documented in detail in the following SOPs:

1) SOP005, "Storage and Transport of Biological Material"
2) SOP038, "Biological Spill Response"

C1.2.4 Local transport out of the laboratory

How will this material be transported on-site (e.g. research material between labs on campus or movement of waste containing viable agents e.g. to a remote autoclave? Detail the containment measures which will be used to prevent or contain accidental splashes or spills

Transfer outside the CBE Laboratory Unit is not anticipated but any requirement is likely to be constrained within the University site. All transport will be subject to controlled procedures according to the local COP and SOP005 (see below). For example, if necessary, transfers will use double containment procedures Transport of research material between laboratories is done using sealed containers which are put into tube racks and trays and transported using trolleys according to the following SOPs. Waste potentially containing viable agents is not removed from the laboratories until it has been autoclaved.

1) SOP003, "Disposal of Biological Waste"
2) SOP005, "Storage and Transport of Biological Material"
3) SOP038, "Biological Spill Response"

C1.2.5 Shipment of Biological Material

Will this material be shipped elsewhere in the UK or abroad?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If yes, give details to support compliance to the relevant regulation (e.g. category of material, correct packaging instruction):	

C1.2.6 Receipt of material

If material will be received from other sites or organisations, what precautions are being taken to ensure that the material is shipped correctly?

All cell lines listed in B2.1.1 are shipped frozen in a dry shipper or double packed by courier. The procedure for the safe receipt of packages containing potentially biohazardous material and their delivery to the appropriate recipient or other designated personnel is documented in SOP008; "Receipt of Hazardous Biological Material". This SOP is intended to minimize the consequences that could result from the failure of packaging methods and materials used to ship biohazardous materials.

C1.2.7 Centrifugation

(i) If material is to be centrifuged will sealed buckets and rotors be used?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes

(ii) Where will these rotors/buckets be opened?

Sealed buckets will be opened within the Containment Level 2 (CL2) Laboratory Unit, unless there is evidence of a potential spillage, in which case the sealed buckets will be opened in the BSC (SOP009, "Use and Maintenance of HERASAFE KS Class II BSC", SOP052, "Use and Maintenance of Bioquell Advanced Microflow Biosafety Cabinet").

The centrifuge is operated and maintained according to the following SOPs:

1) SOP015, "Use and maintenance of BOECO U032R Centrifuge"
2) SOP038, "Biological Spill Response"
3) SOP047, "Use and Maintenance of the Fisher Accuspin Micro-R Centrifuge"

(iii) Describe the procedures in place to deal with leaks and spillages in the centrifuge

Procedures to prevent, contain and respond to leakages and spillages in the centrifuge are detailed in the following SOPs:

1) SOP015, "Use and Maintenance of BOECO U032R Centrifuge"
2) SOP038, "Biological Spill Response"
3) SOP047, "Use and Maintenance of the Fisher Accuspin Micro-R Centrifuge"

Labelled Biological Spill kits are located in each laboratory within the CBE Laboratory Unit. Signs are posted throughout the Laboratory Unit to enable workers to locate the nearest biological (and chemical) spill kits. A biological spill kit is available in Goods Inwards (Wolfson School). Posters are also displayed in each laboratory where a centrifuge is located to advise on spill response and reporting procedures.

C1.2.8 Incubators

If incubators are to be used, what type of incubator (e.g. shaking, static) is used and describe procedures to prevent and contain spillages.

Static incubators are used. Procedures to prevent, contain and respond to spillages in the incubators are detailed in the following SOPs:

1) SOP017, "Use and Maintenance of the Galaxy-R Incubator""
2) SOP053, "Use and Maintenance of Sanyo C02 Incubator"
3) SOP038, "Biological Spill Response"

C1.2.9 Disinfection

Specify the type and concentration of disinfectants to be used:

The disinfectants were carefully chosen for effectiveness in use. The number of disinfectants in use is strictly limited to avoid errors and ambiguities in use and accidental mixing of compounds that may give rise to hazardous reactions or the formation of toxic products. Unless there are compelling reasons to do otherwise, Virkon (1% w/v) is the sole disinfectant used in the laboratories other than 70% IMS which is used for general disinfection cleaning (SOP004) where Virkon cannot be used; for example stainless steel surfaces.

Virkon has a wide range of bactericidal, virucidal, fungicidal and sporocidal activities. Representative viruses from all the major virus families are inactivated by Virkon. Working solutions of 1% w/v have low toxicity and no irritancy. Selection and procedures detailed in the following SOPs:

1) SOP004, "General Laboratory Housekeeping"
2) SOP006, "Selection and Use of Virkon Disinfectant"
3) SOP039, "Storage, Handling and Disposal of Chemicals"

COSHH Risk Assessment reference for Virkon SAF/MM/1745

Have these disinfectants been validated for use with the recipient biological material?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	YES

If yes, describe the procedure:

For Hazard Group 1 and 2 Biological Agents it is normally be sufficient to rely on the manufacturer's data, providing the recommended concentrations and contact times are used. Hence Virkon (1%) is used as per manufacturers instruction and according to standard procedures detailed in the following SOP:

1) SOP006, "Selection and Use of Virkon Disinfectant"

C1.2.10 Personal Protective Equipment (PPE)

(i) What type of lab coats will be worn and where will they be stored? Side fastening Howie type lab coats are worn. They are stored outside the laboratory in purposely designed change rooms. Proper use of PPE is described in the following SOP: SOP037, "Use of Personal Protective Equipment (PPE)"
(ii) What type of gloves will be worn and where will they be stored? 1. Autoclave gloves, which will be stored in close proximity to the autoclave equipment in the Autoclave Room (H31) and the Automated Cell Culture Suite (H21/H22). 2. Cryogenic gloves, which will stored in close proximity to the Liquid Nitrogen storage containers located in Gas Pod 3, Analytical Lab (H23) 3. Latex powder free gloves for general use, which will be stored in the change rooms and point of entry to each laboratory within the CBE Laboratory Unit. Correct use of PPE is described in SOP037, "Use of Personal Protective Equipment (PPE)"
(iii) Describe any other PPE to be used: 1. Laboratory safety glasses (including those for spectacle wearers) 2. Face Shields (primarily for handling liquid nitrogen) 3. Shoe covers, in case of a spillage 4. Aprons or disposable lab coats for extra protection over laboratory coat. Correct use of the above PPE is described in SOP037, "Use of Personal Protective Equipment (PPE)"

C1.2.11 Hygiene Measures

Describe the hygiene facilities available and where they are located 1. Designated hand washing facilities are located in each laboratory change room and in the Analytical Laboratory (H23). 2. Eye Wash stations are located next to each 'hand washing only' sink in each laboratory change room and in the Analytical Laboratory (H23).

C1.2.12 Vaccination

Are effective vaccines available against any of the agents listed in Section1, 2, 3, or 4 of Part B?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe:	

C1.2.13 Waste Treatment before Disposal

How must waste to be treated before disposal and how has it been validated as being effective?		
	Treatment before disposal	Validation
Liquid waste	Virkon sterilise (SOP003 – Disposal of biological waste)	According to manufacturers instructions: see section C2.1.9
Solid waste	Autoclave sterilise (SOP003 – "Disposal of biological waste"	Treatment Cycle validated according to SOP054, " Use and maintenance of the Systec Series 200 Autoclave"

C1.2.14 Autoclave sterilisation

If waste is treated by autoclave sterilisation then this section must be completed. If this section is not relevant then hatch the box			
	Type of waste	*Autoclave cycle (temp, cycle time)*	*Treatment monitor*
Liquid waste			
Solid waste	Cell Culture consumables e.g pipette tips and flasks.	121°C for 15 minutes (under cyclical vacuum)	Designated autoclave tape monitors

Location of autoclave	*Servicing details*	*Location of back-up autoclave*	*Designated area for storage of unsterilised waste*
Automated Cell Culture Suite (H21/22) within the CBE Laboratory Unit i.e. same location as intended work	Annual	Temporary location Lab 208B (Wolfson School) under SOP005, "Storage & Transport of Biological Material"	In secure cage within the Autoclave Room (H31)

C1.2.15 Liquid Waste Disposal

How will liquid waste be disposed of?
To the drain? With copious amounts of water in accordance with SOP003 – " Disposal of biological waste"
As solid waste? No
Other? None

C1.2.16 Solid Waste Disposal

Describe the waste category and disposal route. *(For guidance refer* to http://www.environment-agency.gov.uk)

European Waste Catalogue Code	Categorisation		Disposal Method
		Hatch relevant box(es)	
18 01 01	Sharps		Sharps bin>autoclave sterilisation if known or potentially infected >clinical waste disposal (incineration)
18 01 02 [human]	Human body parts, organs, including blood bags and blood preserves and excreta (unless identified as medium or high risk or known infected in Section 2.16 of this RA in which case they must be pre-treated before disposal and classified 18 01 04 [sealed bins])		Rigid one way sealed tissue bins>incineration only
18 01 02 [animal]	Animal body carcases or recognisable parts ((unless identified as medium or high risk or known infected in Section 2.16 of this RA in which case they must be pre-treated before disposal and classified 18 01 04 [sealed bins])		Rigid one way sealed tissue bins > incineration only

18 01 03	Potentially or known infected lab wastes (including sharps) of HG2, GM Class 2, DEFRA Cat 2 or higher, that have not been pre-treated before leaving the site.		This is not a route of preference and is subject to special requirements
18 01 04 [bags]	Infected or potentially infected lab wastes that have been pre treated before leaving the site		Disinfection or sterilisation (as identified in C1.2.13) in the lab suite > placement in yellow clinical waste bags > clinical waste disposal (incineration)
18 01 04 [sealed bins]	Infected or potentially infected animal or human body parts, organs or excreta that have been pre treated before leaving site		Disinfection or sterilisation (as identified in C1.2.13) in the lab suite > placement in yellow one way sealed tissue bins > incineration)

C1.2.17 Work with Animals or Vectors (if none proceed to Section C1.2.18)

(i) Are animals or vectors to be infected with any of these biological agents?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe the procedure and describe where this aspect of the work will be conducted:	
(ii) Is shedding of infectious materials by the infected animals possible or expected?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe the routes of shedding, risk periods for such shedding and the additional precautions required to control exposure:	
(iii) Who will perform the inoculations of animals/vectors? What training have they received?	
Indicate in the adjacent box if Not Relevant (N/R)	N/R
Provide details of the training required:	

C1.2.18 Bioreactor/Fermenters (if none proceed to Section C1.2.19)

Will a fermenter be used to culture a pathogen?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe the size, and type of the fermenter.	
(ii) Are any supplementary containment measures required, for example, the use of a BSC or spill tray.	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe:	

C1.2.19 Other Control Measures Required?

None

C1.3 Emergency Procedures

C1.3.1 Describe the procedures in place for dealing with spillages *(specify disinfectants and any special containment for large volumes)*

Within the BSC:

Procedures for dealing with small and large spillages are detailed in the following SOPs:

1) SOP006, "Selection and use of Virkon Disinfectant"
2) SOP009, "Use and Maintenance of HERASAFE KS Class II BSC"
3) SOP038, "Biological Spill Response"
4) SOP052, "Use and Maintenance of Bioquell Advanced Microflow Biosafety Cabinet"

Labelled Biological Spill kits are located in each laboratory within the CBE Laboratory Unit. Signs are posted throughout the Laboratory Unit to enable workers to locate the nearest biological (and chemical) spill kits. A biological spill kit is available in Goods Inwards (Wolfson School). Posters are also displayed in each laboratory within the Unit where a BSC is located to advise on spill response (inside the BSC) and reporting procedures.

Within the laboratory but outside the control measure e.g. BSC, spill tray

Procedures for dealing with small and large spillages are detailed in the following SOPs:

1) SOP006, "Selection and use of Virkon Disinfectant"
2) SOP038, "Biological Spill Response"

Labelled Biological Spill kits are located in each laboratory within the CBE Laboratory Unit. Signs are posted throughout the Laboratory Unit to enable workers to locate the nearest biological (and chemical) spill kits. A biological spill kit is available in Goods Inwards (Wolfson School). Posters are also displayed in each laboratory within the Unit to advise on spill response (outside the BSC) and reporting procedures.

Outside the laboratory e.g. during transport

Procedures for dealing with small and large spillages are detailed in the COP and the following SOPs:

1) SOP005, "Storage and Transport of Biological Material"
2) SOP006, "Selection and use of Virkon Disinfectant"
3) SOP038, "Biological Spill Response

Describe the procedures in place for an accidental exposure (if necessary describe different procedures for different types of exposure e.g. eye splash or percutaneous inoculation)

1. Procedures to respond to accidental exposure are detailed in SOP038, "Biological Spill Response" and the local COP. These are detailed in spill response posters located in each laboratory within the Unit. Accident procedures in the case of glass or sharps injury are described in the local COP and displayed in posters located in each laboratory within the Unit
2. Designated hand washing facilities are located in each laboratory change room and in the Analytical Laboratory (H23).
3. Eye Wash stations are located next to each 'hand washing only' sink in each laboratory change room and in the Analytical Laboratory (H23).
4. A First Aid Kit is located outside the Laboratory Unit. Signs are posted throughout the Laboratory Unit to enable workers to locate the nearest Medical Kit. Contact details for First Aiders are posted in each laboratory within the Unit
5. Essential and Emergency Contact details are posted in each laboratory within the Unit.

C2 ASSIGNMENT OF CONTAINMENT LEVEL

The laboratory Containment Level is directly related to each of the 4 Hazard Groups; organisms categorised as HG1 (lowest hazard rating) should normally be handled in CL1 facilities (minimum level of containment), and likewise up to HG4 (highest hazard rating) in CL4 facilities (maximum level of containment). Where the identity or presence of a biological agent is not known the following rules apply: a) where uncertainty exists over the presence of pathogenic biological agent – minimum of CL2; b) where the presence of a pathogenic biological agent is known or suspected – minimum of Containment Level appropriate to the agent, where the assessment is inconclusive but where the activity might involve serious risk – minimum CL3

C2.1. What containment level is required for this work?

All procedures will be carried out under Containment level 2 (CL2) within the CL2 CBE Laboratory Unit. The work activities within this project involve biological agents (BAs) assessed as both Hazard Group 1 and Hazard Group 2. Work with the lower Hazard Group will be carried out under the management standards imposed by the higher level (Containment level 2). This applies under circumstances in which the project is divided into several elements that may be under way in the CBE Laboratory Unit simultaneously. This project, involving the use of Hazard Group 1 BAs that require Containment Level 1 are carried out at Containment Level 2 for reasons other than worker protection; this includes the need to ensure research material protection (e.g. the use of a class II safety cabinet) and to impose a quality assurance discipline

C2.2. Describe extra controls or derogation from certain controls

The CompacT SelecT offers extra controls for automated cell culture processing. The CompacT SelecT (The Automation Partnership, UK) is a fully automated cell culture platform which incorporates a small 6-axis anthropomorphic robotic arm that can access 90 x T175 flask and plate incubators, controlled at 37oC under an atmosphere of 5% CO2 (v/v). Flasks are bar-coded for identification and cell process tracking. Two flask decappers and flask holders, automated media pumping and an automatic cell counter (Cedex®, Innovartis AG, Germany) are integrated within a HEPA filtered cabinet to ensure sterility. The system allows the automation of seeding, feeding and other cell culture processes in order to maintain cell lines in standard T175 cell culture flasks. Risk Assessment reference SAF/MM/1956.

C3 FACILITIES

C3.1 Where will this work take place?

Room(s)	Building	Campus	Person in Control of area
CBE Laboratory Unit *(self contained suite of laboratories and ancillary rooms within the CBE)*	Centre for Biological Engineering	Holywell Park, Loughborough University	Carolyn Thomas Bob Temple Chris Hewitt

C4 PERSONNEL

C4.1 Names of Personnel involved in the Project

Surname	Initials	ID	Position
Chandra	A	5002714	Research Associate
Liu	Y	5003393	Lecturer
Ratcliffe	E	5012183	Research Associate
Rayment	EA	5012060	Research Associate
Thomas	CL	5011765	Laboratory Manager
Thomas	RT	5007730	Lecturer

C4.2 Information, Instruction and Training

Describe the training that will be given to all those affected (directly or indirectly) by the work activity. Instruction should include the 'Local Rules' or 'Local Codes of Practice' which focus on the working instructions to be followed by all persons involved in the work activity to control or prevent exposure to hazardous biological agent(s). These should be written and readily available to all workers working at Containment Level 2. A formal record of training should be kept for all individuals working at Containment Level 2.

Formal records of training are kept for all workers authorised to work at Containment Level 2 (CL2) within the CBE CL2 Laboratory Unit. Instruction against local Code of Practice and QMS i.e. SOPs is provided. Including specific documented training for the Compact Select.

C4.3 Relevant Experience/Training:

Surname	Experience/Training
Chandra A	Documented in Personal Training File
Liu Y	Documented in Personal Training File
Ratcliffe E	Documented in Personal Training File
Rayment EA	Documented in Personal Training File
Thomas CL	Documented in Personal Training File
Thomas RJ	Documented in Personal Training File

C4.4 Other people who may be at risk from the activity e.g. cleaners, maintenance workers or other workers in shared laboratory

Details:

NONE: Cleaners and Maintenance workers are not authorised to enter the laboratory. All laboratory cleaning is undertaken by authorised personnel (i.e. CBE staff). Access for non-laboratory workers is subject to local permit-to-work procedures. If access is needed for essential maintenance of equipment for example a clean down and decontamination of the laboratories will be performed. This will be documented with decontamination certificates and the maintenance worker fully supervised according to SOP004; "General Laboratory Housekeeping" and the local Code of Practice. Two laboratory shut downs occur every year for a week for maintenance work to be done in the CBE Laboratory Unit. Prior to these shut down weeks a full deep clean decontamination will be performed in the all laboratory areas.

All other workers in the CBE Laboratory Unit are authorised personnel.

C5 OCCUPATIONAL HEALTH

C5.1 Vaccination

Is an effective vaccination available for any of the pathogens associated with this work? Advice can be obtained from the Occupational Health Adviser if required. All workers involved with handling unscreened blood, blood products and other tissues are recommended to have Hepatitis B immunization

Certificate for Hepatitis B Immunization documented in personal training file

C5.2 Health Surveillance

Is health surveillance required? (Health surveillance is typically applied if working with a hazardous substance that: a) produces an identifiable disease or adverse health effect that can be related to exposure; b) there is a reasonable likelihood that the disease or effect may occur under the conditions of work, and c) there are valid techniques for detecting indications of the disease or effect).

None required

C6. NOTIFICATIONS: Human Tissue Act

C6.1.1 Relevant material covered by the Human Tissue Act

Are any of the cell, tissues or fluids to be used covered by the Human Tissue Act?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No

C6.1.2 Does This Work Have Ethical Approval? If Yes, Provide Details

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes
Approval number: 08/H0406/122	
Date obtained: 19.08.08	Ethics committee name: NHS Research Ethics Committee: Leicestershire, Northampton & Rutland EC1

C6.1.3 Are other registrations/notifications required for this work? For example HSE notification under COSHH, Home Office notification under anti-terrorism, crime and security act etc

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If Yes, give details:	

7. LICENSING REQUIREMENTS FOR ANIMAL PRODUCTS

C7.1.1 Are there any licensing requirements for this work?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
NOTE: The regulations covering the import of animal products (including tissue cultures, tissues, body fluids or fractions thereof) are in a state of flux. See the DEFRA website for details. UNLESS THIS SECTION IS NOT RELEVANT (N/R) ie THE INTENDED WORK DOES NOT USE ANIMAL PRODUCTS - CONSULT THE LU H&S OFFICE TO REVIEW APPLICATION REQUIREMENTS BEFORE ANY SUBMISSIONS	

8. DECLARATION

*The declaration must be signed **before** submitting this assessment to the Departmental Safety Officer and University Biological Safety Officer*

I, the undersigned:

- confirm that all information contained in this assessment is correct and up to date
- will ensure that **suitable and sufficient instruction, information and supervision** is provided for all individuals working on the activity
- will ensure that no work will be carried out until this **assessment has been completed and approved** and that all necessary control measures are in place
- accept that for some Containment Level 2 and all CL3 activities **a statutory notification period of 20 days** may be required before work can commence
- that all information contained in this assessment must remain correct and up to date (the assessment should be **reviewed once a year** and whenever any **significant changes** to the work activity occur)
- will re-submit the assessment for approval if any significant changes occur

Name: Person conducting assessment	Signature	Date
Carolyn Thomas	cthomas	11 May 2009.
Paul Hourd	P Hourd	11 MAY 2009
Name: Principal Investigator	**Signature**	**Date**
David J Williams		12 May 09

9.APPROVAL		
Name: Departmental Safety Officer	**Signature**	**Date**
C.J. Hewett		11/V/09
Name: University Biological Safety Officer	**Signature**	**Date**
C. MOORE	C. M. Moore.	27/5/09.

CBE/BRA/008

FORM LU/HE/BRA

Version 1.0 Oct 08

RISK ASSESSMENT OF WORK WITH BIOLOGICAL MATERIALS

Please note the following before completing this form:

1. University Health and Safety Policy requires that risk assessment of all work with biological materials must be carried out in advance of work commencing. A key requirement of The Control of Substances Hazardous to Health Regulations (COSHH) is to assess the risks associated with any work activity involving the use of biological materials that may contain biological agents.
2. YOU SHOULD COMPLETE ALL OF PART A, THE APPROPRIATE SECTION(S) OF PART B, AND ALL OF PART C. THIS FORM SHOULD BE SUBMITTED TO THE HEALTH AND SAFETY UNIT FOR REVIEW (VIA YOUR DEPARTMENTAL SAFETY OFFICER)
3. It is the responsibility of the Principal Investigator to ensure compliance to these requirements and that this risk assessment remains valid.
4. This risk assessment form **IS NOT** for assessing the risks associated with **Genetically Modified Organism activities.**

Date Submitted:		Date Approved:	

PART A: Please provide the following general information:

School/Department			
Healthcare Engineering, Wolfson School of Mechanical & Manufacturing Engineering			
The Project			
Title of Project: REMEDI – Grand Challenge – WP3.(ii) This Risk Assessment applies to the final one and a half years of the aforementioned project			
Project Reference Number: REMEDI/WP3(ii)			
Person responsible for this work (*Principle Investigator*):			
Name: David Williams:		Position: Professor of Healthcare Engineering	
Department: Healthcare Engineering		University School: Wolfson School of Mechanical and Manufacturing Engineering	
Person conducting this assessment			
Name: Carolyn Thomas/Paul Hourd		Position: Laboratory Manager	
Department:	Healthcare Engineering	Date Risk Assessment Undertaken:	October 2008
Proposed Project Start Date:	Nov 2008	Proposed Project End Date:	28.02.2010

Assessment Review:
required at least once a year or immediately following any significant change to the project

	Review 1	Review 2	Review 3	Review 4
Due Date				
Date Conducted				

A1 PROJECT SUMMARY

A1.1 Scientific Goals of the Project *Brief yet clear outline only*

Cells- Process Discovery, Optimisation and continuous improvement for the manufacture of cellular feedstocks.

1) The development and demonstration of the application of designated experiments for the establishment of process conditions- process discovery and optimisation-for the volume processing of complex human cell types of commercial importance in tissue engineering at commercially relevant scales. The development of statistical models of the process under a variety of conditions.

2) The development and laboratory demonstration of the use of continuous improvement techniques to improve the process output in the manufacture of tissue engineered components and systems that are capable of being compliant with current and emerging regulation.

3) Analysis of the effectiveness of the chosen approach, and generation of methods for improving the resource effectiveness of the process optimisation approach

(As documented in Remedi Grand Challenge proposal)
(jeSRPI)

A1.2 Description of the Experimental Procedures

Describe laboratory procedures to be used and highlight any non-standard laboratory operations

1) Human osteoblast-like cell line (HOS) – Seeded onto tissue culture treated plastic (e.g. T75 flasks) and cultured at 37°C 5% CO^2 until 70% confluent, followed by trypsinisation and subsequent culture. Manual and automated cell culture techniques will be used.

2) Human mesenchymal progenitor cells (hMSCs)- Isolated via centrifugation of fresh bone marrow aspirate over a defined a defined density cell separation product (Histopaque™). Cells will be seeded onto tissue culture treated plastic (e.g. T175 flasks) and cultured at 37°C 5% CO^2 until 70% confluent, followed by trypsinisation and subsequent subculture .Manual and automated cell culture techniques will be used . Cryopreservation and subsequent thawing and subculture will be conducted at various points in the process.

3)Human mesenchymal stem cells from Bone Marrow mononuclear cells (hMSCs)- Cells will be seeded onto tissue culture treated plastic (e.g. T175 flasks) and cultured at 37°C 5% CO^2 until 70% confluent, followed by trypsinisation and subsequent subculture. Manual and automated cell culture techniques will be used. Cryopreservation and subsequent thawing and subculture will be conducted at various points in the process.

4)Endothelial Progenitor from peripheral blood mononuclear cells- (derived from Poietics® mononuclear cells from Human G-CSF Mobilised cells), Cells will be cultured on human fibronectin coated T175 flasks and the adherent cells passaged according to the general culture method.

5) Human Embryonic cell line (hES)-(Hues-1,Hues-9) - hES cells will be cultured in treated plastic vessels (e.g. T175 flasks) and cultured at 37°C 5% CO^2, followed by enzyme digestion and subsequent subculture. Cryopreservation and subsequent thawing and subculture will be conducted at various points in the process.. Manual and automated cell culture techniques will be used.

6) Mouse Embryonic Fibroblasts (MEFs) – MEF cells obtained in 15 T175 flasks from the University of Nottingham will be cultured at 37°C 5% CO^2 . Media in the cells will be changed every 24h hours, pooled and frozen in 100ml Nalgene sterile media bottles for subsequent hES culture. Various manual and automated culture routines will be used.

All procedures will be conducted in accordance with the laboratory Quality Management System (QMS) requirements, Good Cell Culture Practice, Good Aseptic Technique and the local and University Codes of Practice (COP).

PART B: Please provide information in one or more of the following sections, as appropriate. Only sections which you complete should be submitted:

Section 1: *micro-organisms (prions, viruses, bacteria, fungi, parasites in ACDP category 2 and pathogens controlled by the Department for the Environment, Food and Rural Affairs).* ***[Work with ACDP category 3 and 4 pathogens is not currently permitted in the University.]***

Section 2: *cell cultures, tissues, blood, body fluids or excreta*

Section 3: *plants and plant material*

Section 4: *animals and animal tissues*

SECTION 2: CELL CULTURES, TISSUES, BLOOD, BODY FLUIDS OR EXCRETA

B2.1 HAZARD AND RISK IDENTIFICATION: NATURE OF CELLS, TISSUES OR BODY FLUIDS

*This information gives an indication of the **potential** harm that the biological material may cause*

B2.1.1 List all cells or tissues to be used. *For cells indicate if primary, continuous or finite.*

Indicate in the adjacent box if Not Relevant (N/R)			
Cell or tissue type and ID	Organ Source	Species	From where will it be obtained?
1) Human Osteoblast like cell line (HOS)	Bone	Human	European Collection of Cell Culture (ECACC), UK Continuous cell line
2)Human Mesenchymal progenitor cells (hMSCs)	Bone Marrow	Human	Cambrex,UK Primary cell line
3) Human Mesenchymal stem cells from Bone marrow mononuclear cells	Bone Marrow	Human	Lonza,UK Primary Cell line
4)Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF mobilised cells	Blood	Human	Lonza,UK Primary Cell line
5) Human Embryonic cell line (hES) (Hues-1,)	Embryo	Human	Harvard University,USA Continuous cell line
6) Human Embryonic cell line (hES) (Hues-9)	Embryo	Human	Harvard University,USA Continuous cell line
7) Mouse Embryonic fibroblasts (MEFs)	Skin	Mouse	Nottingham University,UK Continuous cell line

B2.1.2 List all blood, body fluids or excreta to be used

Indicate in the adjacent box if not relevant (N/R			N/R
Material type and ID	Organ Source	Species	From where will it be obtained?

B2.1.3 Has any material listed in section B2.1.1 been genetically modified in any way?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If Yes, complete Genetically Modified Organisms (GMO) Risk Assessment Form	

B2.1.4 Will material be screened for infectious agents (if from a cell culture collection answer B2.1.6)?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	YES

If Yes, provide details of the types of screening and agents screened for:

Cell or tissue type	Screened for
1) Human osteoblast-like cell line (HOS)	See section B2.1.6. Cultured cells in-house tested for mycoplasma
2) Human Mesenchymal progenitor cells (hMSCs)	Basic chemistry panel, complete blood count, HIV-I, Hepatitis B and C, PRP, Urinalysis, sickle cell solubility test and pregnancy test of donors by the supplier. Refer to Cambrex product data sheet*
3) Human Mesenchymal stem cells from bone marrow mononuclear cells	HIV-I, Hepatitis B and C testing by supplier. Refer to Lonza product data sheet*
4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF Mobilised cells	Each donor is tested and found non-reactive by an FDA approved method for the presence of HIV-1, Hepatitis B Virus and Hepatitis C Virus by the supplier. Refer to Lonza Material Safety Data Sheet*. Cultured cells in-house tested for mycoplasma
5) Human Embryonic cell line (hES) (Hues-1,)	Academic source. No material safety data sheet available See B2.1.6
6) Human Embryonic cell line (hES) (Hues-9,)	Academic source. No material safety data sheet available See B2.1.6
7) Mouse Embryonic Fibroblasts (MEFs)	Academic source. No material safety data sheet available

** (All Material Safety Data Sheets will be held in a folder in the Healthcare Engineering Research Office)*

B2.1.5 Will any clinical history (if relevant) be provided with this material?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	Yes

If yes give details:

(2) Human Mesenchymal progenitor cells (hMSCs) – A data sheet is provided with the cells containing information on sample collection and clinical history of donors.

(4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF Mobilised cells - A data sheet is provided with the cells containing information on sample collection and clinical history of donors.

If yes, will a policy of rejection of samples from diseased patients be adopted? Explain

Not relevant for materials (2) and (4) because diseased patient samples are not procured by the commercial supplier/vendor. The data sheets provide information on donor eligibility criteria.

If yes, how will the information be disseminated in the course of the project?

Not relevant for materials (2) and (4) – donor eligibility is determined by pre-screening tests carried out by the supplier as above.

If yes, will this information be anonomised?

For materials (2) and (4), donor information received from the commercial supplier/vendor is anonymised

B2.1.6 If obtained from a cell culture collection, is safety information provided?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	Yes

If Yes, summarise here:

Two of the agents listed in B2.1.4 are obtained from a Cell Culture Collection

1) Human osteoblast-like cell line – There is no evidence for the presence of infections, virus or toxic products. However, ECACC recommends that cultures are handled in Category 2 containment. Refer to ECACC material data sheet.

2) Human embryonic cell line (hES)- (Hues-1,Hues-9) – Obtained from the Howard Hughes Medical Institute, Harvard University (originator) solely for internal academic research purposes. The material is experimental in nature and may have hazardous properties since not all of its characteristics are known. See reference Material Use Licence F191, Harvard University office of Technology Development.

B2.2 RISK TO HUMANS

B2.2.1 What is the likelihood of infection of this material? Indicate as None, Low Risk, Medium Risk, High Risk, Known Infected*

Cell type and ID	Risk Category	Justification for Selection
1) Human osteoblast-like cell line (HOS)	Low	Well authenticated/characterised continuous cell line from a culture collection. Has documented provenance and screened for the most serious human pathogens. Screened as described in section B2.1.6. Baseline containment level CL1
2)Human Mesenchymal progenitor cell (hMSCs)	Low	Well authenticated/characterised primary cell line from a commercial source. Has documented provenance and screened for the most serious human pathogens. Screened as described in section B2.1.4. Baseline containment level CL2
3) Human Mesenchymal stem cell from Bone marrow mononuclear cell.(hMSCs)	Low	Well authenticated/characterised primary cell line from a commercial source. Has documented provenance and screened for the most serious human pathogens. Screened as described in section B2.1.4. Baseline containment level CL2
4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF mobilised cells	Low	Well authenticated/characterised primary cell line from a commercial source culture collection. Has documented provenance and screened for the most serious human pathogens. Screened as described in section B2.1.4. Baseline containment level CL1
5) Human embryonic cell (hES) (Hues-1,)	Low	Well authenticated continuous cell line obtained from the originators culture collection The cells are not fully characterised but have been utilised extensively in peer reviewed academic research. These cells are sourced from a leading academic laboratory that has successfully deposited cells to the National Institute for Biological Standards and Control stem cell bank. Since these cell lines have been subject to extensive sub-culture the risk of pathogenic agent contamination is very low. Baseline containment level CL2
6) Human embryonic cell (hES) (Hues-9)	Low	Well authenticated continuous cell line obtained from the originators culture collection The cells are not fully characterised but have

		been utilised extensively in peer reviewed academic research. These cells are sourced from a leading academic laboratory that has successfully deposited cells to the National Institute for Biological Standards and Control stem cell bank. Since these cell lines have been subject to extensive sub-culture the risk of pathogenic agent contamination is very low. Baseline containment level CL2
7) Mouse Embryonic Fibroblasts (MEFs)	Low	Not fully characterised continuous cell line of animal origin obtained from the academic originator, as described in section B2.1.4. Baseline containment level CL1
		If low risk or none proceed to section B2.2.4

see The Managing the risks in laboratories and healthcare premises – available at
http://www.hse.gov.uk/biosafety/biologagents.pdf

B2.2.2 If medium or high risk (section B2.2.1), name and classify the Biological Agents this material could be infected with. List the biological agents and indicate the ACDP hazard group classification*

Name of Agent	Classification

see The Approved List of Biological Agents – available on the Health & Safety website or
http://www.hse.gov.uk/pubns/misc208.pdf.

B2.2.3 Describe the routes of infection (in humans) for these adventitious agents (place a 'X' in the relevant box)

Cell or Tissue type	Percutaneous	Mucocutaneous	Inhalation	Ingestion
Details:				

B2.2.4 Are there any other biological hazards (other than adventitious infectious risk) associated with the materials e.g. tumourogenic cells

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If Yes, describe:	

B2.3 HUMANS AT INCREASED RISK OF INFECTION

B2.3.1 Do any of the agents listed in section 2.1 present an overt risk to humans at increased risk (including immunocompromised workers, pregnant workers, breast feeding mothers)?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, Occupational Health must be consulted:	

B2.4. PROPAGATION OR CONCENTRATION OF ADVENTITIOUS AGENTS

B2.4.1 Will any culturing of this material take place?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	Yes
If yes, identify the cells and the conditions these will grow: All cells listed are cultured using manual and automated techniques (see section A1.2) in tissue culture flasks in cell culture media in a 37 degree Celsius humidified incubator.	

B2.4.2 If culturing, will CD4+ cells be present. Describe what cells and for how long these cultures will be allowed to grow

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, explain:	

B2.4.3 If culturing, what is the maximum volume of culture grown?

Indicate in the adjacent box if Not Relevant (N/R)		
	Per Flask	Per experiment
1) Human osteoblast-like cell line (HOS)	$6x10^6$ (75ml flask)	Variable- depends on experiment
2) Human Mesenchymal progenitor cells (hMSCs)	~10 million	~ 200 million (20 flasks)
3) Human Mesenchymal stem cell from Bone Marrow mononuclear cells (hMSCs)	~10 million	~ 200 million (20 flasks)
4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF mobilised cells	~10 million	~ 200 million (20 flasks)
5) Human embryonic cell line (hES) (Hues-1)	~10 million	~ 200 million (20 flasks)
6) Human embryonic cell line (hES) (Hues-9)	~10 million	~ 200 million (20 flasks)
7) Mouse Embryonic Fibroblasts (MEFs)	~10 million	~ 200 million (20 flasks)

B2.4.4 Will the cells be manipulated in any way that could result in a concentration of any adventitious biological agent present?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, explain:	

B2.5 WORKING WITH MATERIAL DONATED BY YOURSELF OR COLLEAGUES : Persons MUST NOT work with their own cells.

B2.5.1 Will any cells be donated by persons working in or has access to the lab?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, explain what precautions are to be taken to prevent that person being exposed to the cells:	
If yes, where will this material be collected:	
If yes, provide justification for not using a safer source:	
If yes, how will confidentiality be assured:	
If yes, has Ethics Committee approval been obtained:	

B2.6 ENVIRONMENTAL CONSIDERATIONS:

B2.6.1 Are any of the agents capable of causing disease or other harm in animals, fish or plants?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, describe:	

B2.6.2 Will there be any other environmental risks?

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	No
If yes, describe:	

B2.7 OTHER HAZARDS

B2.7.1 Are there any other hazards associated with this work? For example, hazardous chemicals, cryogenic gases ionising radiation.

Indicate in the adjacent box as: Yes, No or Not Relevant (N/R)	Yes
If yes, identify these: 1) Cryogenic processing with Liquid Nitrogen 2) Trypan Blue for generic cell viability testing 3) Generic and specific hazardous non-biological materials eg Histopaque etc.	
If yes, have these been risk assessed and any necessary approval obtained? 1) Liquid Nitrogen.- Procedures will be carried out by trained personnel in accordance with SOP013 "Use and Maintenance of Liquid Nitrogen Stores". Risk Assessment Reference: SAF/MM/1638 2) Trypan Blue - Procedures will be carried out by trained personnel in accordance with SOP029 " Handling and disposal of Trypan blue". Risk (COSHH) Assessment Reference SAF/MM/1745 3) All Hazardous non-biological materials used in this project eg Trypan Blue, Histopaque etc are subjected to COSHH assessment.	

SECTION 4: ANIMALS AND ANIMAL TISSUES

B4.1 HAZARD AND RISK IDENTIFICATION: NATURE OF ANIMALS OR TISSUE

*This information gives an indication of the **potential** harm that the biological material may cause*

B4.1.1 List all animals or animal tissues to be used

Species	Sex	Source	Anatomical Site	Origin or geographical source
Foetal Calf Serum (FCS)	Unknown	Bovine Foetus	Foetus	Commercial supplier: Cambrex, UK. Sourced from South America according to data sheet
Matrigel ™ (mouse)	Unknown	Engelbreth-Holm-Swarm mouse Sarcoma	Basement Membrane matrix mouse tumour	Commercial supplier: Becton Dickinson, (BD), Oxford, UK

B4.1.2 Is the animal or tissue/body fluid to be worked with infected or to be infected?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No

If Yes, complete Section 1 of this form

FCS product contains material of animal origin. The material contains no hazardous or toxic substances. The material is supplied by a commercial company and sent through the post in secure packaging. Material is liquid and stored frozen at -20C.

Matrigel product is derived from mouse colonies that are routinely screened for pathogens. It is tested and found negative for bacteria, fungi and mycoplasma by the supplier. The material is supplied by a commercial company and sent through the post in secure packaging. Material is liquid and stored frozen at -20C.

B4.1.3 Is a carcinogen, drug or other substance to be administered to the animal(s) or present in the tissue?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R

If Yes, complete the appropriate Chemical COSHH Assessment

B4.1.4 Have the investigators that will be performing the work on animals obtained the appropriate Home Office Licence?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R

If No, consult the H&S Office.

B4.1.5 Have Standard Operating Procedures (SOPs) for the proposed work been approved?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R

If No, consult the H&S Office. If Yes attach the signed approval.

B4.2 RISK TO HUMANS

B4.2.1 The disease(s) caused to humans
Describe the type and severity of effects or disease(s) on human health (including infection, allergy, bites and scratches)

Name of animal/animal tissue	Type	Severity
Foetal Calf Serum	Under EC regulations (Directive 1999/45/EC), contains no hazardous or toxic substances according to the supplier. Likelihood that it contains substances hazardous to health is low. Animal proteins may be a potential contact irritant	Potential contact irritant
Matrigel ™	Product contains no hazardous constituents, or concentration of all chemical constituents are below the regulatory threshold limits described by Occupational Safety Health Hazard Communication Standard 29CFR 1910.1200 and the EU Directive 91/155/EEC and 93/112/EC. Refer to MSDS*	N/R

B4.2.2 What is the likelihood of infection of this material? INDICATE as None, Low Risk, Medium Risk, High Risk, Known Infected

Name of agent	Risk Category	Justification for Selection
Non categorised (refer to MSDS)	None.	FCS product contains no hazardous or toxic substances that requires Cambrex to distribute a MSDS according to EC regulations. Well authenticated/characterised product from commercial source. Baseline containment level CL1
Non categorised (refer to MSDS)	None	Matrigel™ is used in small quantities for research use only. Well authenticated/characterised product from commercial source. Baseline containment level CL1. Mouse colonies are routinely screened for pathogens via Mouse antibody Production (MAP). Tested and found negative for bacteria, fungi and mycoplasma by the supplier. Refer to Matrigel™ Material Safety Data Sheet*
		If none proceed to section B4.3

B4.2.3 Describe the routes that the effects described in section B4.2.1 are transmitted (place a 'X' in the relevant box)

Percutaneous	Mucocutaneous	Inhalation	Ingestion	N/R

Details:

B4.3 HUMANS AT INCREASED RISK OF INFECTION

B4.3.1 Do any of the agents listed in section B4.1 present an overt risk to humans at increased risk (including immunocompromised workers, pregnant workers, breast feeding mothers, workers repeatedly handling or multiply dosing animals)?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If yes, Occupational Health must be consulted:	

B4.4. PROPAGATION OR CONCENTRATION OF ADVENTITIOUS AGENTS

B4.4.1 Will any culturing of this material take place?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If yes, complete Section 2 of this form:	

B4.4.2 How many animals will be used?

Indicate in the adjacent box if Not Relevant (N/R)	N/R

B4.5 ENVIRONMENTAL CONSIDERATIONS: Risk to other animals

B4.5.1 Will there be any risk other animals?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe:	

B4.5.2 Will there be any other environmental risks?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If yes, describe:	

PART C: CONTROL MEASURES

C1. CONTROL MEASURES

The risk of exposure must be prevented or adequately controlled to minimise the chance of harm arising. COSHH Regulations require minimum containment measures for laboratories handling organisms from the different ACDP hazard groups (http://www.hse.gov.uk/pubns/misc208.pdf)
The hazard group number typically indicates the level of containment (includes physical measures & working practices) that must be used for its handling).

C1.1 Preventing Exposure

C1.1.1 Substitution with a Safer Alternative

Is substitution with a safer alternative practical, by for example, replacement of a clinical strain or pathogen with one that is lab adapted? Provide reasons for your answer:

Cell /Tissue Type	Substitution available ?
1) Human osteoblast-like cell line (HOS)	Not available. Use of these specific human cells is critical to the value of the research. Cells are sourced from established commercial suppliers according to SOP048 " Generation of Risk Assessments for New Materials and Processes."
2 Human Mesenchymal progenitor cell (hMSCs)	
3) Human Mesenchymal stem cells from bone marrow mononuclear cells (hMSCs) -	
4) Endothelial Progenitor from peripheral blood Poietics® mononuclear cells from human G-CSF Mobilised cells	
5) Human Embroyonic cell line (Hues -1)	Not available. Use of these specific embryonic cells is critical to the value of the research. Cells are sourced from an established culture collection at Harvard University.
6) Human Embroyonic cell line (Hues -9)	
7) Mouse Embryonic fibroblasts (MEFs)	Not available. Use of these specific cells is critical to the value of the research. These cells are sourced from Nottingham University and their specific use designated by the transferred protocol for eSC culture.
8) Foetal Calf Serum (FCS)	Not available. Use is needed for supplementing media for cell culture. Sourced from established commercial supplier according to SOP048 " Generation of Risk Assessments for New Materials and Processes"
9) Matrigel™	Not Available. Use designated by specific transferred culture protocols. Sourced from established commercial suppliers.

C1.1.2 Isolation/Segregation

(i) Is/Are the laboratory(s) to be used for this work to be shared with other workers not directly involved in this activity?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes

If yes, provide details:

Access restricted to authorised lab workers with appropriate training in accordance with documented Code of Practice (COP) and Quality Management System (QMS) requirements for Containment Level 2 work (CL2).

The laboratories are locked at all times when not in use to ensure safe storage of biological agents. Keys to the labs are only issued to authorised users.

(ii) Is access to the laboratory(s) to be used for this work restricted?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes
If yes, provide details: Restricted to people with appropriate training (authorised access documented in individual training records) in accordance with the COP and QMS	

C1.2 Controlling Exposure

C1.2.1 Are sharps (needles, blades, scissors, forceps, glass or capillary tubes) to be used at any stage during this activity?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If yes, list the sharps:	
If yes, justify there use – is there an alternative?:	
If yes, describe there use and disposal:	
If yes, describe any additional precautions employed to reduce risk:	

C1.2.2 Containment and Ventilation

(i) Is the use of BSC required for the protection of the worker ie do the work procedures generate aerosols or splashes that pose a risk to workers?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes
If yes, specify the type(s) and when they will be used: A Class II Biological Safety Cabinet will be used for all manipulations according to the following SOPs 1) SOP009, "Use and Maintenance of BSC-G2000 Vertical Laminar Airflow Cabinet" 2) SOP052, "Use and Maintenance of Bioquell Class II Cabinet" 3) SOP035, "Use and Maintenance of CompacT SelecT" The Compact Select is used to seed, feed, maintain, expand and harvest human cells (primary and cell lines) cultured in bar-coded T175 conventional flasks. Both multiple individual flask cultures and multiple flask batches are maintained concurrently and processed without cross contamination, mix-ups or other detrimental effects. The Compact Select is the equivalent of a BSC and incubator in one unit, therefore containing any harmful aerosols and splashes inside the equipment; see section C2.2. Risk Assessment for the Compact Select SAF/MM/1956	
(ii) Are there any requirements for room ventilation e.g. negative pressure, temperature control?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If yes, specify:	

C1.2.3 Transport and Storage within the laboratory

How and where are materials to be stored?

All cell lines/tissues listed in B2.1.1 will be stored in a cryobank or temporary storage in designated cell culture incubators according to the following SOPs :

1) SOP005, "Storage and Transport of Biological Agents"
2) SOP013, "Use and maintenance of Liquid Nitrogen Stores"
3) SOP017, "Use and maintenance of the Galaxy-R Incubator"
4) SOP031, "Cryopreservation and Storage of Mammalian Cell Lines"
5) SOP053, "Use and maintenance of the Sanyo CO2 Incubator

Foetal Calf Serum and Matrigel will be stored in Fridges and Freezers according to the following SOP's:

1) SOP016 " Use and Maintenance of fridges and freezers"
2) SOP005 " Storage and Transport of Biological Agents "
3) SOP039 " Storage, Handling and disposal of Chemicals"

How will this material be transported within the laboratory e.g. between BSC and incubator? Detail the containment measures which will be used to prevent or contain accidental splashes or spills.

Cells will always be transferred in closed secondary containers large enough to carry the designated material. . Appropriate spill response procedures are posted in the lab and documented in detail in the following SOPs:

1) SOP005, "Storage and Transport of Biological Agents"
2) SOP038, "Biological Spill Response"

C1.2.4 Local transport out of the laboratory

How will this material be transported on-site (e.g. research material between labs on campus or movement of waste containing viable agents e.g. to a remote autoclave? Detail the containment measures which will be used to prevent or contain accidental splashes or spills

Transfer outside the laboratory is not anticipated but any requirement is likely to be constrained within the Wolfson building . If necessary, transfers will use double containment procedures Transport of research material between laboratories is done using sealed containers which are put into tube racks and trays and transported using trolleys according to the following SOPs. Waste potentially containing viable agents is not removed from the laboratories until it has been autoclaved.

1) SOP003, "Disposal and Disinfection of Biological Waste"
2) SOP005, "Storage and Transport of Biological Agents"
3) SOP038, "Biological Spill Response"

C1.2.5 Shipment of Biological Material

Will this material be shipped elsewhere in the UK or abroad?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No
If yes, give details to support compliance to the relevant regulation (e.g. category of material, correct packaging instruction):	

C1.2.6 Receipt of material

If material will be received from other sites or organisations, what precautions are being taken to ensure that the material is shipped correctly?

All cell lines listed in B2.1.1 are shipped frozen in a dry shipper or double packed by courier. The procedure for the safe receipt of packages containing potentially biohazardous material and their delivery to the appropriate recipient or other designated personnel is documented in SOP008; "Receipt of Purchased Biohazardous Material". This SOP is intended to minimize the consequences that could result from the failure of packaging methods and materials used to ship biohazardous materials.

C1.2.7 Centrifugation

(i) If material is to be centrifuged will sealed buckets and rotors be used?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes

(ii) Where will these rotors/buckets be opened?

Sealed buckets will be opened within the Containment Level 2 (CL2) laboratory, unless there is evidence of a potential spillage, in which case the sealed buckets will be opened in the BSC (SOP009, "Use and Maintenance of BSC-G2000 Vertical Laminar Airflow Cabinet", SOP052, "Use and Maintenance of Bioquell Class II Cabinet").

The centrifuge is operated and maintained according to the following SOPs:

1) SOP015, "Use and maintenance of BOECO U032R Centrifuge"
2) SOP038, "Biological Spill Response"

(iii) Describe the procedures in place to deal with leaks and spillages in the centrifuge

Procedures to prevent, contain and respond to leakages and spillages in the centrifuge are detailed in the following SOPs:

1) SOP015, "Use and maintenance of BOECO U032R Centrifuge"
2) SOP038, "Biological Spill Response"

The spill kit is located under the sink in the laboratory Foyer. A second Spill Kit is available in Goods Inwards Posters are also displayed around the laboratory to advise on spillages.

C1.2.8 Incubators

If incubators are to be used, what type of incubator (e.g. shaking, static) is used and describe procedures to prevent and contain spillages.

Static incubators are used. Procedures to prevent, contain and respond to spillages in the incubators are detailed in the following SOPs:

1) SOP017, "Use and maintenance of the Galaxy-R Incubator""
2) SOP053, "Use and maintenance of Sanyo C02 Incubator"
3) SOP038, "Biological Spill Response"

C1.2.9 Disinfection

Specify the type and concentration of disinfectants to be used:

The disinfectants were carefully chosen for effectiveness in use. The number of disinfectants in use is strictly limited to avoid errors and ambiguities in use and accidental mixing of compounds that may give rise to hazardous reactions or the formation of toxic products. Unless there are compelling reasons to do otherwise, Virkon (1% w/v) is the sole disinfectant used in the laboratories other than 70% IMS which is used for general disinfection cleaning (SOP004) where Virkon cannot be used; for example stainless steel surfaces.

Virkon has a wide range of bactericidal, virucidal, fungicidal and sporocidal activities. Representative viruses from all the major virus families are inactivated by Virkon. Working solutions of 1% w/v have low toxicity and no irritancy.

Selection and procedures detailed in the following SOPs:
1) SOP004 General Laboratory Maintenance and cleaning
2) SOP006 Selection and Use of Disinfectants
3) SOP039 Storage, Handling and Disposal of Chemicals

COSHH Risk Assessment reference for Virkon SAF/MM/1745

Have these disinfectants been validated for use with the recipient biological material?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes

If yes, describe the procedure:

For Hazard Group 1 and 2 Biological agents it is normally be sufficient to rely on the manufacturers data, providing the recommended concentrations and contact times are used. Hence Virkon (1%) is used as per manufacturers instruction and according to standard procedures detailed in the following SOP:
1) SOP006, "Selection and Use of Disinfectants

C1.2.10 Personal Protective Equipment (PPE)

(i) What type of lab coats will be worn and where will they be stored?

Side fastening lab coats are worn which have elasticated cuffs. They are stored outside the laboratory.. Proper use of PPE is described in the following SOP: SOP037, "Use of Personal Protective Equipment"

(ii) What type of gloves will be worn and where will they be stored?

1. Autoclave gloves, which will be stored in close proximity to the autoclave equipment
2. Cryogenic gloves, which will stored in close proximity to the Liquid Nitrogen storage containers
3. Latex powder free gloves for general use, which will be stored in the laboratory

Correct use of PPE is described in SOP037, "Use of Personal Protective Equipment"

(iii) Describe any other PPE to be used:

1. Laboratory safety glasses (including those for spectacle wearers)
2. Face Shields (primarily for handling liquid nitrogen)
3. Shoe covers, in case of a spillage
4. Aprons or disposable lab coats for extra protection over laboratory coat.

Correct use of the above PPE is described in SOP037, "Use of Personal Protective Equipment"

C1.2.11 Hygiene Measures

Describe the hygiene facilities available and where they are located

1) Eye Wash station located in the laboratory foyer
2) Hand washing facilities located in the laboratory foyer

C1.2.12 Vaccination

Are effective vaccines available against any of the agents listed in Section1, 2, 3, or 4 of Part B?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R

If yes, describe:

C1.2.13 Waste Treatment before Disposal

How must waste to be treated before disposal and how has it been validated as being effective?		
	Treatment before disposal	Validation
Liquid waste	Virkon sterilise (SOP003 – disposal and disinfection of biological waste)	According to manufacturers instructions: see section C2.1.9
Solid waste	Autoclave sterilise (SOP003 – "Disposal and disinfection of biological waste"	Treatment Cycle validated according to SOP010, " Use and maintenance of Boxer autoclave"

C1.2.14 Autoclave sterilisation

If waste is treated by autoclave sterilisation then this section must be completed. If this section is not relevant then hatch the box			
	Type of waste	*Autoclave cycle (temp, cycle time)*	*Treatment monitor*
Liquid waste			
Solid waste	Cell Culture consumables e.g pipette tips and flasks.	121°C for 1 hour	Designated autoclave tape monitors

Location of autoclave	*Servicing details*	*Location of back-up autoclave*	*Designated area for storage of unsterilised waste*
Laboratory T208B i.e. same location as intended work	Annual	Lab 207	On designated benches adjacent to the autoclave

C1.2.15 Liquid Waste Disposal

How will liquid waste be disposed of?
To the drain? With copious amounts of water in accordance with SOP003 – " Disposal and disinfection of biological waste"
As solid waste? No
Other? None

C1.2.16 Solid Waste Disposal

Describe the waste category and disposal route. *(For guidance refer* to http://www.environment-agency.gov.uk)

European Waste Catalogue Code	Categorisation		Disposal Method
		Hatch relevant box(es)	
18 01 01	Sharps		Sharps bin>autoclave sterilisation if known or potentially infected >clinical waste disposal (incineration)

18 01 02 [human]	Human body parts, organs, including blood bags and blood preserves and excreta (unless identified as medium or high risk or known infected in Section 2.16 of this RA in which case they must be pre-treated before disposal and classified 18 01 04 [sealed bins])		Rigid one way sealed tissue bins>incineration only
18 01 02 [animal]	Animal body carcases or recognisable parts ((unless identified as medium or high risk or known infected in Section 2.16 of this RA in which case they must be pre-treated before disposal and classified 18 01 04 [sealed bins])		Rigid one way sealed tissue bins > incineration only
18 01 03	Potentially or known infected lab wastes (including sharps) of HG2, GM Class 2, DEFRA Cat 2 or higher, that have not been pre-treated before leaving the site.		This is not a route of preference and is subject to special requirements
18 01 04 [bags]	Infected or potentially infected lab wastes that have been pre treated before leaving the site		Disinfection or sterilisation (as identified in C1.2.13) in the lab suite > placement in yellow clinical waste bags > clinical waste disposal (incineration)
18 01 04 [sealed bins]	Infected or potentially infected animal or human body parts, organs or excreta that have been pre treated before leaving site		Disinfection or sterilisation (as identified in C1.2.13) in the lab suite > placement in yellow one way sealed tissue bins > incineration)

C1.2.17 Work with Animals or Vectors (if none proceed to Section C1.2.18)

(i) Are animals or vectors to be infected with any of these biological agents?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe the procedure and describe where this aspect of the work will be conducted:	
(ii) Is shedding of infectious materials by the infected animals possible or expected?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe the routes of shedding, risk periods for such shedding and the additional precautions required to control exposure:	
(iii) Who will perform the inoculations of animals/vectors? What training have they received?	
Indicate in the adjacent box if Not Relevant (N/R)	N/R
Provide details of the training required:	

C1.2.18 Bioreactor/Fermenters (if none proceed to Section C1.2.19)

Will a fermenter be used to culture a pathogen?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe the size, and type of the fermenter.	
(ii) Are any supplementary containment measures required, for example, the use of a BSC or spill tray.	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If yes, describe:	

C1.2.19 Other Control Measures Required?

None

C1.3 Emergency Procedures

C1.3.1 Describe the procedures in place for dealing with spillages *(specify disinfectants and any special containment for large volumes)*

Within the BSC: Procedures for dealing with small and large spillages are detailed in the following SOPs: 1) SOP006, "Selection and use of Disinfectants" 2) SOP009, "Use and Maintenance of BSC-G2000 Vertical Laminar Airflow Cabinet" 3) SOP038, "Biological Spill Response" 4) SOP052, "Use and Maintenance of Bioquell Class II Cabinet" A spill kit is located under the sink in the laboratory Foyer. A second Spill Kit is available in 'Goods Inwards' Posters displayed within the laboratory detail what to do in the event of a spillage.
Within the laboratory but outside the control measure e.g. BSC, spill tray Procedures for dealing with small and large spillages are detailed in the following SOPs: 1) SOP006, "Selection and use of Disinfectants" 2) SOP038, "Biological Spill Response" A spill kit is located under the sink in the laboratory Foyer. A second Spill Kit is available in 'Goods Inwards'
Outside the laboratory e.g. during transport Procedures for dealing with small and large spillages are detailed in the COP and the following SOPs: 1) SOP005, "Storage and Transport of Biological Agents" 2) SOP006, "Selection and use of Disinfectants" 3) SOP038, "Biological Spill Response The spill kit is located under the sink in the laboratory Foyer. A second Spill Kit is available in 'Goods Inwards'
Describe the procedures in place for an accidental exposure (if necessary describe different procedures for different types of exposure e.g. eye splash or percutaneous inoculation) Procedures to respond to accidental exposure are detailed in the following SOP: 1) SOP038, "Biological Spill Response" Handwashing facilities, an eye wash station, First Aid Kit and contact details for First Aiders are available in the laboratory Foyer.

C2 ASSIGNMENT OF CONTAINMENT LEVEL

The laboratory Containment Level is directly related to each of the 4 Hazard Groups; organisms categorised as HG1 (lowest hazard rating) should normally be handled in CL1 facilities (minimum level of containment), and likewise up to HG4 (highest hazard rating) in CL4 facilities (maximum level of containment). Where the identity or presence of a biological agent is not known the following rules apply: a) where uncertainty exists over the

presence of pathogenic biological agent – minimum of CL2; b) where the presence of a pathogenic biological agent is known or suspected – minimum of Containment Level appropriate to the agent, where the assessment is inconclusive but where the activity might involve serious risk – minimum CL3

C2.1. What containment level is required for this work?

All procedures will be carried out under Containment level 2 (CL2).

C2.2. Describe extra controls or derogation from certain controls

The CompacT SelecT offers extra controls for automated cell culture processing. The CompacT SelecT (The Automation Partnership, UK) is a fully automated cell culture platform which incorporates a small 6-axis anthropomorphic robotic arm that can access 90 x T175 flask and plate incubators, controlled at 37oC under an atmosphere of 5% CO2 (v/v). Flasks are bar-coded for identification and cell process tracking. Two flask decappers and flask holders, automated media pumping and an automatic cell counter (Cedex®, Innovartis AG, Germany) are integrated within a HEPA filtered cabinet to ensure sterility. The system allows the automation of seeding, feeding and other cell culture processes in order to maintain cell lines in standard T175 cell culture flasks. Risk Assessment reference SAF/MM/1956.

C3 FACILITIES

C3.1 Where will this work take place?

Room(s)	Building	Campus	Person in Control of area
T208B T207	Wolfson School of Mechanical &Manufacturing Engineering	Loughborough University	Carolyn Thomas Bob Temple

C4 PERSONNEL

C4.1 Names of Personnel involved in the Project

Surname	Initials	ID	Position
Chandra	A	5002714	Research Associate
Liu	Y	5003393	Lecturer
Ratcliffe	E	5012183	Research Associate
Rayment	EA	5012060	Research Associate
Thomas	CL	5011765	Laboratory Manager
Thomas	RJ	5007730	Lecturer

C4.2 Information, Instruction and Training

Describe the training that will be given to all those affected (directly or indirectly) by the work activity. Instruction should include the 'Local Rules' or 'Local Codes of Practice' which focus on the working instructions to be followed by all persons involved in the work activity to control or prevent exposure to hazardous biological agent(s). These should be written and readily available to all workers working at Containment Level 2. A formal record of training should be kept for all individuals working at Containment Level 2.

Formal records of training are kept for all workers at Containment Level 2 (CL2). Instruction against local QMS ie SOPs and the local COP is provided. Including Specific training for the Compact Select

C4.3 Relevant Experience/Training:

Surname	Experience/Training
Chandra A	Documented in Personal Training File
Liu Y	Documented in Personal Training File
Ratcliffe E	Documented in Personal Training File
Rayment EA	Documented in Personal Training File
Thomas CL	Documented in Personal Training File
Thomas RJ	Documented in Personal Training File

C4.4 Other people who may be at risk from the activity e.g. cleaners, maintenance workers or other workers in shared laboratory

Details:

None:

Cleaners and Maintenance workers are not authorised to enter the laboratory. If access is needed for essential maintenance of equipment a clean down and decontamination of the laboratories will be performed. This will be documented with decontamination certificates and the maintenance worker fully supervised according to SOP004 " General Laboratory Maintenance and cleaning" Two laboratory shut downs occur every year for a week for maintenance work to be done in the laboratory. Prior to these shut down weeks a full deep clean decontamination will be performed in the laboratory areas.

Other authorised workers may be in the laboratory.

C5 OCCUPATIONAL HEALTH

C5.1 Vaccination

Is an effective vaccination available for any of the pathogens associated with this work? Advice can be obtained from the Occupational Health Adviser if required. All workers involved with handling unscreened blood, blood products and other tissues are recommended to have Hepatitis B immunization

Certificate for Hepatitis B Immunization documented in personal training file

C5.2 Health Surveillance

Is health surveillance required? (Health surveillance is typically applied if working with a hazardous substance that:

a) produces an identifiable disease or adverse health effect that can be related to exposure; b) there is a reasonable likelihood that the disease or effect may occur under the conditions of work, and c) there are valid techniques for detecting indications of the disease or effect).

No

C6. NOTIFICATIONS: Human Tissue Act

C6.1.1 Relevant material covered by the Human Tissue Act

Are any of the cell, tissues or fluids to be used covered by the Human Tissue Act?	
Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	No

C6.1.2 Does This Work Have Ethical Approval? If Yes, Provide Details

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	Yes
Approval number: 08/H0406/122	
Date obtained: 19.08.08	Ethics committee name: NHS Research Ethics Committee: Leicestershire, Northampton & Rutland EC1

C6.1.3 Are other registrations/notifications required for this work? For example HSE notification under COSHH, Home Office notification under anti-terrorism, crime and security act etc

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R
If Yes, give details:	

7. LICENSING REQUIREMENTS FOR ANIMAL PRODUCTS

C7.1.1 Are there any licensing requirements for this work?

Indicate in the adjacent box as No, Yes or Not Relevant (N/R)	N/R

NOTE: The regulations covering the import of animal products (including tissue cultures, tissues, body fluids or fractions thereof) are in a state of flux. See the DEFRA website for details.

UNLESS THIS SECTION IS NOT RELEVANT (N/R) ie THE INTENDED WORK DOES NOT USE ANIMAL PRODUCTS - CONSULT THE LU H&S OFFICE TO REVIEW APPLICATION REQUIREMENTS BEFORE ANY SUBMISSIONS

8. DECLARATION

*The declaration must be signed **before** submitting this assessment to the Departmental Safety Officer and University Biological Safety Officer*

I, the undersigned:

- confirm that all information contained in this assessment is correct and up to date
- will ensure that **suitable and sufficient instruction, information and supervision** is provided for all

individuals working on the activity

- will ensure that no work will be carried out until this **assessment has been completed and approved** and that all necessary control measures are in place
- accept that for some Containment Level 2 and all CL3 activities **a statutory notification period of 20 days** may be required before work can commence
- that all information contained in this assessment must remain correct and up to date (the assessment should be **reviewed once a year** and whenever any **significant changes** to the work activity occur)
- will re-submit the assessment for approval if any significant changes occur

Name: Person conducting assessment	**Signature**	**Date**
Carolyn Thomas	cethomas	31/10/08
Paul Hourd	PHourd	31/10/08
Name: Principal Investigator	**Signature**	**Date**
D.T. WILLIAMS		3/11/08

9.APPROVAL		
Name: Departmental Safety Officer	**Signature**	**Date**
R I Tomalo		09/11/08
Name: University Biological Safety Officer	**Signature**	**Date**